

重离子束辐射技术在花卉育种中的应用

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摘 要: 重离子束辐射是一种新兴的辐射诱变技术。与传统的 γ 射线、X射线等相比, 重离子束具有高传能线密度, 能够产生更高的相对生物学效应, 具有诱变率更高、诱变范围更大的特点。常用的离子源主要有 C、N、Ne 等, 不同植物材料的不同部位和发育阶段对重离子束的敏感度效果不一。通过结合植物组织培养、辐射材料处理、多次辐射和基因工程等技术, 目前已对菊花、香石竹、美女樱和夏堇等 20 多种植物开展了重离子束辐射育种研究, 获得了新种质和新品种, 部分新品种在市场推广后获得了良好的经济效益。因此, 重离子束辐射技术在观赏植物育种上有着广泛的应用前景。

关键词: 重离子束; 辐射; 观赏植物; 育种

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Application of Heavy Ion Beam Irradiation in Ornamental Flowers Breeding

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Abstract: Heavy ion beam irradiation is an emerging radiation mutagenesis technology. Compared with traditional radiation such as X-rays and gamma rays, heavy ion beam provides a high linear energy transfer (LET) and relative biological effectiveness (RBE). This technique induces mutations at a higher rate and wider mutation spectrum. When irradiated, the sensitivity of different parts and developmental stages of different plant materials varies. Radiation ion source commonly used is C, N, Ne etc. Through a combination of tissue culture, materials processing, re-irradiation and genetic engineering, this approach has been applied in more than 20 ornamental plants, including chrysanthemum, carnation, verbena, torenia etc. Many novel mutations and varieties have been obtained, some of them have been commercialized with large economic benefits. Heavy ion beam technology is an excellent tool to improve mutation breeding of horticulture plants with high efficiency.

Key words: heavy ion beam; irradiation; ornamental plant; breeding

近年来, 重离子束辐射 (heavy-ion beam irradiation) 作为一种新兴的辐射诱变源越来越受到关注。重离子是指荷能质量数大于 4 的带电粒子 (周利斌 等, 2008), 具有高的传能线密度 (带电粒

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子在单位长度径迹上传递的能量, linear energy transfer, LET) 和相对生物学效应 (relative biological effectiveness, RBE), 其单位剂量的诱变效应高于 X 射线、 γ 射线、电子束等 (Okamura et al., 2003; Shikazono et al., 2003; 周利斌 等, 2008)。重离子通过加速器注入生物体内, 同生物材料中的分子、原子发生一系列的碰撞 (王菊芳 等, 2007), 能在局部区域释放出高能量, 显著诱导单链或双链 DNA 断裂、末端受损, 从而不容易修复 (Goodhead, 1995)。

重离子束辐射是一种很有效的诱导技术手段, 可以有效的改变目标性状, 能在不影响其他特征的基础上提供一个广泛的变异范围 (Shikazono et al., 2005), 通过 γ 射线辐射未能获得的特殊花色可以通过离子辐射育种诱导出 (Okamura et al., 2003), 从而提高园艺和农作物的诱变频率。在对拟南芥 (*Arabidopsis thaliana*) 的碳离子束辐照研究中发现, 一半的变异为点变异, 另一半有着大的 DNA 改变, 例如插入、易位和大片段缺失 (Shikazono et al., 2005)。一般认为重离子束能产生比较大的 DNA 变换, 插入、易位和大片段缺失, 而不是点变异 (Tanaka & Hase, 2009)。目前开展重离子束辐射育种的单位及其设备主要有日本理化学研究所的放射性同位素束流设施 (Riken RI Beam Factory); 日本原子能机构的高崎量子应用研究所 (Takasaki Ion Accelerators for Advanced Radiation Application); 日本若狭湾能源研究中心的多用途同步串联加速器 (Wakasa Wan Energy Research Center Multi-purpose Accelerator with Synchrotron and Tandem); 中国科学院近代物理研究所 (Institute of Modern physics) 和意大利国家核物理研究所的南方国家实验室 (Laboratori Nazionali del Sud) 等。

1 辐射材料与方法

不同的植物材料对辐射的敏感性有很大差异, 表现在不同物种、品种以及同一植株的不同发育阶段和器官上 (Kazama et al., 2008; 孙兰弟, 2008)。目前用于重离子束辐射的材料有干种子、吸水饱和种子、芽、叶片、花瓣、愈伤组织、组培小苗、体细胞胚等, 它们对重离子束的敏感度为: 愈伤组织 > 发育中胚 > 茎节 > 吸水饱和种子 > 干种子 (Abe et al., 2007, 2012; Kazama et al., 2008)。

一般说来, 组培苗具有较强的分化和形成新组织、器官的能力, 是辐射育种较理想的材料 (彭绿春 等, 2007); 且植物组织培养用的外植体材料含水量较干种子要高得多, 辐射敏感性相应也高得多 (周利斌 等, 2007)。但也有一些例外, 例如在仙客来 (*Cyclamen persicum*) 中, 比起愈伤组织、体细胞胚等, 用重离子束辐射块茎更能有效的改变花色性状 (Sugiyama et al., 2008)。在菊花 (*Chrysanthemum morifolium*) 中, 和叶片相比, 辐射花瓣后结合组织培养能获得诱变率更高的花色突变体, 推测花色相关基因在花瓣中表达更为活跃, 从而更容易发生变异 (Hase et al., 2009)。

进行辐射所用的离子源主要有 C、N、O、Ne、Ar、Fe 等。目前的研究结果显示, 离子束辐射诱变的影响并非取决于离子源的种类, 而是取决于它的传能线密度 LET。目前应用较多的离子源是碳离子, 常用 LET 值的范围为 $100 \sim 200 \text{ keV} \cdot \mu\text{m}^{-1}$ (Atsushi & Yoshihiro, 2010)。中国科学院近代物理研究所用低能氮、碳离子束辐照万寿菊 (*Tagetes erecta*)、一串红 (*Salvia splendens*)、矮牵牛 (*Petunia hybrida*)、长春花 (*Catharanthus roseus*)、鸡冠花 (*Celosia cristata*) 的种子和大丽花 (*Dahlia pinnata*) 的花芽, 获得了观赏性状改变的诱变植株 (卫增泉 等, 2003; 周利斌 等, 2008)。

2 对观赏植物的诱变

对观赏植物而言花色花型是重要的观赏性状, 通过进行重离子束辐射, 诱导出了很多新的表型。例如获得了复杂花色和条纹花色的菊花以及腋芽减少的菊花突变体, 并对该诱变植株进行了再次辐

射，获得了腋芽减少且在低温下能够开花的诱变植株（Atsushi & Yoshihiro，2010）。重离子束辐射后的香石竹（*Dianthus caryophyllus*）植株表现出了比 X 射线辐照更多的花色变异种类（Okamura et al., 2003）。通过重离子束诱导还获得了花朵粉色、樱桃红色，有条纹和斑点的矮牵牛植株（Okamura et al., 2009）。在月季（*Rosa*）中，获得了花色更加艳丽、花瓣数量、花的大小和花形都发生改变的诱变植株（Yamaguchi et al., 2003）。在夏堇（*Torenia fournieri*）中，通过离子束辐射获得了两类花色变异体，一种是缺乏花色合成的基因，另一种是花色基因的表达发生了改变（Miyazaki et al., 2006）。

同时，对许多观赏植物而言，不育性状是很重要的。一方面能够增加花朵数量和延长花期，另一方面雄性不育的植株不会通过授粉将其基因传递到自然环境中（Sugiyama et al., 2008）。美女樱（*Verbena hybrida*）经过重离子束辐射后成功的分离出不育变异体 ‘Temari Bright Pink’，是世界上第 1 个通过重离子束辐射技术获得的新品种，在 2002 年春季进入商业市场（Kazama et al., 2008）。此外，由于辐射诱变的不确定性，还获得了一些株形、植株高度、叶色等性状发生变异的突变体。目前开展重离子辐射的植物材料及结果见表 1。

表 1 重离子束辐射观赏植物获得的变异后代
Table 1 Mutant lines developed in various ornamentals using heavy ion beam breeding

序号 No.	植物材料 Plants species	辐射部位 Irradiated materials	离子束种类和参数 Ion-beam and physical parameters	获得突变体 Mutation induction	参考文献 References
1	矮牵牛 <i>Petunia hybrida</i> ‘Kirin Hana-Saka Rose’	离体芽 <i>In vitro</i> buds	$^{12}\text{C}^{6+}$ (320 MeV)	花色粉和樱桃红、花瓣中细白条纹和 斑点的变异植株 Flower color mutants with pink, cherry, minute striped white, and spotted petals	Okamura et al., 2009
2	矮牵牛 <i>Pentunia hybrida</i> ‘BBss11’	播种小苗 Seedlings	$^{12}\text{C}^{6+}$ (320 MeV)	花色洋红、紫红、淡粉、白、蓝色白 边和深紫红等变异植株 Mutants of magenta, purple, light pink, white, blue picotee and burgundy flowers	Hase et al., 2009, 2010
3	巴西水竹叶 <i>Tradescantia fluminensis</i>	嫩枝生长点 Growing points of shoots	^{12}C (95.8 MeV · u ⁻¹)	彩叶、花葶紫、花萼白、花朵淡粉的 变异植株 Mutant with color leaves, purple stalk, chlorophyll-deficient calyces and light pink flower	He et al., 2011
4	大丽花 <i>Dahlia</i> ‘Miharu’	嫩枝 Shoots	^{14}N (135 MeV · u ⁻¹)	花色深粉、浅粉、粉白等和花径变异 的诱变植株 Flower color mutants (dark pink, pale pink and pink-white etc.) and flower diameter mutants	Abe et al., 2005
5	大丽花 <i>Dahlia</i> spp.	组培苗生长点 Tissue-cultured shoots	$^{12}\text{C}^{6+}$ (320 MeV)	花色、花形变异的诱变植株 Mutants on flower color and flower shape	Uyama et al., 2012
6	飞燕草 <i>Delphinium</i> ‘Monoka’	叶片 Leaf	$^{12}\text{C}^{6+}$ (320 MeV)	花色淡粉、红粉、黄粉，花序小花增 多和矮生的变异植株 Flower color mutants (light pink, red pink and yellow pink), mutant with increased number of spikes and dwarf mutant	Chinone et al., 2009; Kitamura et al., 2010
7	鹤石斛 <i>Dendrobium crumenatum</i>	原球茎 Protocorm-like bodies	$^{12}\text{C}^{6+}$ (320 MeV)	花朵变大、花葶变长的变异植株 Mutants with large flowers, a longer flower stalk	Affrida et al., 2008
8	宫灯百合 <i>Sandersonia aurantiaca</i>	干种子 Dry seeds	^{14}N (28.5 keV · μm ⁻¹)	白化变异植株 Albino mutant plants	Horita et al., 2002; Mohan Jain, 2006; Abe et al., 2012
9	菊花 <i>Chrysanthemum morifolium</i> ‘Reagan Red’	舌状花 Ray floret	$^{12}\text{C}^{6+}$ (320 MeV)	株高、花形、花色、花朵大小和数量， 叶色和叶形等突变 Mutants on plant height, flower shape, flower color, number of flower bud and flower size, leaf color and shape	Affrida et al., 2010; Shakinah et al., 2011

续表 1

序号 No.	植物材料 Plants species	辐射部位 Irradiated materials	离子束种类和参数 Ion-beam and physical parameters	获得突变体 Mutation induction	参考文献 References
10	菊花 <i>Chrysanthemum</i> 'Jimba'	叶片 Leaves	$^{12}\text{C}^{5+}$ (220 MeV); $^{12}\text{C}^{6+}$ (320 MeV)	低温下开花和腋芽减少的变异植株 Mutants with few axillary buds, low temperature flowering trait	Ueno et al., 2003
11	菊花 <i>Chrysanthemum</i> 'Jimba' 腋 芽减少的诱变植株 'Aladdin' few axillary buds mutant 'Aladdin' induced from <i>Chrysanthemum</i> 'Jimba'	叶片 Leaves	$^{12}\text{C}^{5+}$ (220 MeV); $^{12}\text{C}^{6+}$ (320 MeV)	腋芽减少且在低温下能够开花的变 异植株 Mutants with few axillary flower buds and low temperature flowering	Ueno et al., 2004
12	菊花 <i>Chrysanthemum</i> 'Hakusui'	离体试管芽 <i>In vitro</i> buds	$^{12}\text{C}^{6+}$ (320 MeV)	不定芽减少和低温下开花的变异植 株 Mutants with less adventitious buds, no flowering delay under low temperature condition	Asami et al., 2010; Hase et al., 2011
13	农杆菌介导的转 <i>pac1</i> 基因菊花 <i>Dendranthema grandiflora</i> agrobacterium-mediated transgenic <i>chrysanthemum</i>	叶片 Leaf segments	$^{12}\text{C}^{6+}$ (320 MeV)	舌状花浅粉、深粉、浅橙、白和黄色 等变异植株 Pale pink, dark pink, salmon, white and yellow etc. ray florets mutants	Okamura et al., 2008
14	菊花 <i>Chrysanthemum morifolium</i> 'H13'	叶片 Leaf segments	$^{12}\text{C}^{5+}$ (220 MeV)	舌状花深红、朱红、粉红、红白、红 白重瓣, 和 T 型花簇的变异植株 Floret color mutants (dark-red , vermilion, pink, white-red, white-red double blooms), and mutant with T type of flower cluster	Furutani et al., 2008; Matsumura et al., 2010
15	菊花 <i>Chrysanthemum morifolium</i> 'Shiroyamate'	舌状花 Ray florets	$^{12}\text{C}^{5+}$ (220 MeV)	舌状花黄和淡黄的变异植株 Yellow and pale yellow ray florets mutants	Matsumura et al., 2010
16	菊花 <i>Chrysanthemum morifolium</i> 'Taihei'	茎节的腋芽 Nodes with axillary buds	^{12}C (220 MeV)	花朵浅粉、深粉、白、黄、浅黄、橙、 深橙、红等的变异植株 Mutants with pale pink, deep pink, white, yellow, pale yellow, orange, deep orange and red flowers	Yamaguchi et al., 2009
17	蓝眼菊 <i>Osteospermum</i> 'Mother Symphony'	叶片 Leaf segments	$^{12}\text{C}^{5+}$ (220 MeV); $^{12}\text{C}^{6+}$ (320 MeV)	花色白、奶油、深黄和橙, 斑叶和矮 生的变异植株 Flower color mutants (white, cream, deep yellow and orange), dwarf mutants and variegated leaf mutants	Iizuka et al., 2006
18	蓝眼菊 <i>Osteospermum</i> 'Mother Symphony' 经离子辐射后的诱变 株 OM7 OM7 induced from radiated mutants <i>Osteospermum</i> 'Mother Symphony'	叶片 Leaf section	$^{12}\text{C}^{5+}$ (220 MeV); $^{12}\text{C}^{6+}$ (320 MeV)	花色白紫、淡橙、和花瓣背面黄色的 变异植株 White purple and light bitter orange flower mutants , and mutant with light yellow in the back side of the petal	Iizuka et al., 2008
19	蓝眼菊诱变株 <i>Osteospermum</i> 'Mother Symphony' OM7 再次离 子辐射后诱变株 OM706 OM706 induced from re-irradiated mutants <i>Osteospermum</i> 'Mother Symphony' OM7	叶片 Leaf sections	$^{12}\text{C}^{5+}$ (220 MeV); $^{12}\text{C}^{6+}$ (320 MeV)	花朵黄白、浅橙、浅黄、淡黄色和花 背面亮黄色的变异植株 Flower color mutants with yellowish white, whitish light orange, whitish light yellow, and lighter yellow in the back side of the petal	Okada et al., 2009
20	毛油点草 <i>Tricyrtis hirta</i>	花被片诱导的 胚性愈伤组织 Embryogenic calluses induced from tepals	$^{12}\text{C}^{6+}$ (135 MeV · u ⁻¹)	矮生、叶形、花朵大小和数量变异的 植株 Mutants on dwarf, leaf shape, flower number and flower size	Nakano et al., 2010a
21	美女樱 <i>Verbena</i> × <i>hybrida</i> 'Temari Sakura', 'Temari Coral Pink'	含有侧生分生 组织的茎节 Nodes containing lateral meristems	$^{14}\text{N}^{7+}$ (1.89 GeV)	花数量增多、花期变长、不育的变异 植株 Mutants with a large number of inflorescences, better longevity, and sterile trait	Kanaya et al., 2008

续表 1

序号 No.	植物材料 Plants species	辐射部位 Irradiated materials	离子束种类和参数 Ion-beam and physical parameters	获得突变体 Mutation induction	参考文献 References
22	天竺葵 <i>Pelargonium</i> ‘Splendide’	茎节上的腋芽 Stem node containing a single axillary bud	^{12}C (23 keV · μm^{-1})	雄蕊萎缩、无花粉的雄性不育诱变植株 Male sterile mutants lacked pollen-producing ability	Sugiyama et al., 2005; Mohan Jain, 2006; Abe et al., 2012
23	夏堇 <i>Torenia</i> ‘Summer Wave Blue’	叶组织和不含侧生分生组织的茎节 Leaf tissue and stem internodes without lateral meristems	^{14}N (135 MeV · u^{-1}); ^{20}Ne (135 MeV · u^{-1})	花朵淡蓝、蓝、淡粉和粉色的变异植株 Pale blue, blue, pale pink and pink flower mutants	Miyazaki et al., 2006
24	夏堇和转基因 (<i>CHS</i> 和 <i>DFR</i>) 株系 <i>Torenia fournieri</i> ‘Crown Violet’ and transgenic lines	组培苗叶片 Leaf disks from <i>in vitro</i> plants	^{12}C (1.62 GeV); ^{20}Ne (2.70 GeV)	花色更柔和、镶边、条纹等和花瓣圆形、波状、流苏状等变异植株 Flower color and coloration pattern mutants (tone-shifted, bordered, streaked etc.), petal shape and corolla divergence mutants (rounded, wavy, fringed etc.)	Sasaki et al., 2008
25	仙客来 <i>Cyclamen persicum</i> ‘Fragrance Mini’	体胚小苗的块茎 Tubers of plantlets induced from somatic embryos	$^{12}\text{C}^{6+}$ (1.62 GeV)	花色淡蓝紫色加白色条纹、淡红紫色等和花瓣形状变异的变异嵌合体及雄性不育植株 Chimeric mutants on flower color (reddish purple, striped pattern of white-purple) and petal forming, and male sterile mutants	Sugiyama et al., 2008
26	仙客来 <i>Cyclamen persicum</i> × <i>C. purpurascens</i> ‘Kaori-no-mai’	种子组培暗培养后的黄化叶柄 Etiolated petioles induced from germinated seedlings (kept in dark)	$^{12}\text{C}^{6+}$ (320 MeV)	花红紫色的变异植株 Red-purple flowers mutant	Kondo et al., 2009, 2010
27	香石竹 <i>Dianthus caryophyllus</i> ‘Vital’	叶片 Leaf segments	$^{12}\text{C}^{5+}$ (220 MeV)	花红、深粉、粉、浅粉、黄、双色、条纹等和花型石竹型、月季型等的变异植株 Flower color mutants (red, dark pink, pink, light pink, yellow, bi-colored, striped) and flower shape mutants (Dianthus type and rose type)	Okamura et al., 2003
28	一串红 <i>Salvia splendens</i>	干种子 Dry seeds	$^{12}\text{C}^{6+}$ (80.55 MeV · u^{-1})	深红色和鲜红色花朵变异植株 Dark red and fresh red flower color mutants	Wu et al., 2009
29	月季 <i>Rosa</i> ‘Orange Rosamini’	组培苗的茎节 Stems <i>in vitro</i> culture	^{12}C (220 MeV)	花色更加鲜艳、浅红色、花瓣数量、花朵大小、和花型发生变异的突变体 Mutants on flower color (intense flower color and reddish flower), petals number, flower size and flower shape	Yamaguchi et al., 2003
30	非洲紫罗兰 <i>Saintpaulia ionantha</i> ‘Mauve’, ‘Indikon’	幼嫩叶片 Young leaves	$^{12}\text{C}^{6+}$ (960 MeV)	获得了叶绿素缺失突变体 Chlorophyll deficiency mutants	Zhou et al., 2006

3 与其他技术的结合

为了提高目的性状的诱变频率和稳定性,常需要将重离子束辐射技术与其他技术相结合。

一是组织培养技术。辐射诱变育种过程中,突变嵌合体的分离和稳定变异体的获得是关键内容之一(彭绿春等,2007)。重离子束辐射结合组织培养能够更有效的对突变体进行诱导和筛选,将其中的有益突变固定和增殖,有效的加速了新品种培育(Okamura et al., 2003; Zhou et al., 2006; Sugiyama et al., 2008; Matsumura et al., 2010; Nakano et al., 2010b)。同时由于花卉主要是靠无性繁殖方式进行扩繁,重离子辐照结合组织培养为花卉产业提供了新的育种途径(周利斌等,2008),在培育园艺新品种上有着广阔的应用前景。

二是辐射材料处理技术。对辐射材料采取恰当的预处理能够有效提高目标性状的诱变频率。例如发现用高浓度蔗糖溶液对矮牵牛和萝卜小苗进行预处理,可以显著提高色素的积累,提高花色变异植株的诱变率(Hara et al., 2003; Hase et al., 2010)。

三是重复重离子束辐射技术。对突变体进行离子再辐射以获得丰富的变异性状(Yamaguchi et al., 2003; Iizuka et al., 2008)。例如为了获得纯白色舌状花的蓝眼菊,对诱变植株进行多次重离子束辐射(Okada et al., 2009)。

四是基因工程技术。通过重离子束辐射技术获得新的诱变植株后,可以通过基因工程技术获得新基因。例如通过重离子束辐照拟南芥后获得了多种突变体,从抗紫外线B突变体 *uvi1* 和 *uvi4* 分离鉴定了相关基因(Tanaka et al., 2002; Hase et al., 2006);从花青素积累 *ast* 突变体或花青素缺乏 *tt19* 突变体中获得了部分相关基因(Tanaka et al., 1997; Kitamura et al., 2004);从锯齿状花瓣和萼片的突变体 *frill1* 获得了与该表型相关的基因等(Hase et al., 2000, 2005)。

4 讨论与展望

同常规辐射育种相比,重离子束辐射育种有着诱变效率更高,诱变谱更广的特点。结合植物无性繁殖技术,对辐射后的材料进行栽培、选育和突变体鉴定等,获得具有优良性状的新品种,缩短了育种年限。然而重离子束辐射育种中也存在着一些缺点。例如诱变率不高,仍需要在大量诱变植株中进行选育;目的性较差,辐射突变方向不定(程金水,2000);以及重复性不高,在不同的试验中获得相同的变异植株较为困难(Kondo et al., 2008; Sasaki et al., 2008)。此外,辐射剂量的多少除了对植物的诱变效果有所影响外,也对植物生长发育有所影响,例如碳离子束辐射石斛(*Dendrobium mirbellianum*)后生长十分缓慢(Affrida et al., 2008);以及一串红的花色突变体在所有的生长阶段都受到抑制,但在开花和结实上没有任何影响(Wu et al., 2009)。经重离子束辐射后,毛油点草的胚性愈伤组织的生长也受到了抑制,但在低剂量辐射时明显促进了愈伤组织中体细胞胚的生长(Nakano et al., 2010b)。因此一般选择相对较低的辐射剂量,既有较高的诱变率,又不会对植物生长有明显抑制。总体而言,重离子束辐射在花卉新品种培育方面有着很好的研究、开发与应用前景。

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